

# THE EL10 SUGAR BEET GENOME (AND BEYOND)

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**Mitch McGrath**, Paul Galewski, Andy Funk, Belinda Townsend, Karen Davenport, Hajnalka Daligault, Shannon Johnson, Joyce Lee, Alex Hastie, Aude Darracq, Glenda Willems, Steve Barnes, Ivan Laichko, Shawn Sullivan, Sergey Koren, Adam Phillippy, Jie Wang, Tiffany Lu, Jane Pulman, Kevin Childs, Anastasia Yocum, Damian Fermin, Shujun Ou, Piergiorgio Stevanato, Kazunori Taguchi

[mitchmcg@msu.edu](mailto:mitchmcg@msu.edu)

## Why assemble the EL10 sugar beet genome?

If you have one genome, you have one genome (*differences matter*).

Improved and 'affordable' technology for  
(a more) *contiguous & accurate* genome assembly.

Chromosomes resolved to *single nucleotide* level (~ smallest unit of change).

An interesting technical challenge.

Progenitor of EL10 is parent to >23,000 hybrids and inbred derivatives,  
>6,700 of these are inbred 6 to 9 generations (RILs).

# Sequencing the EL10 beet: inputs

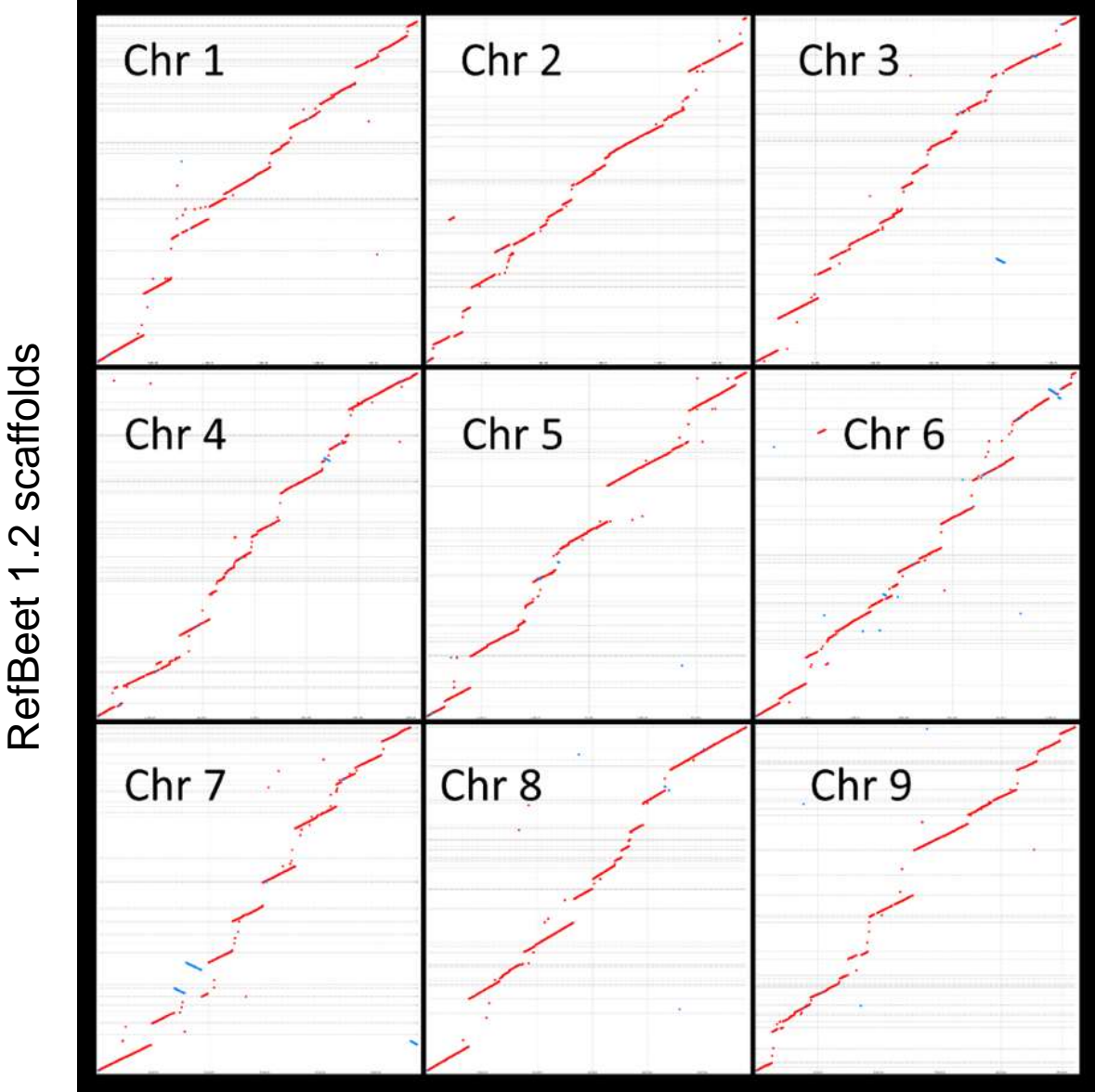
Technology	Library		Coverage <sup>1</sup>
		<b>PacBio passed reads</b>	
PacBio long reads	RS II, P6-C4 chemistry (Los Alamos Nat'l Labs)	6,540,795	79.3
	Mean length = 9,096 nt (std.dev = 6,528)		
	> 40 kb initial mapping and pre-assembly	5,176	0.38
		<b>BioNano passed labels</b>	
Optical physical map	BioNano Genomics, <i>Bss</i> SI - <i>Bsp</i> PQ1 Hybrid Scaffold	121 Gb	161.3
	<i>Bsp</i> PQ1 (7.6 labels/100 kb)	40 Gb	
	<i>Bss</i> SI (10 labels/100 kb)	81 Gb	
		<b>Illumina passed reads</b>	
Paired-End short reads	HiSeq 2500, TruSeq Libraries, 125bp PE (MSU-RTSF)	447,211,041	149
Cross-linked <i>in vivo</i>	Phase Genomics Hi-C library, HiSeq 2500, TruSeq Libraries	355,892,798	118.6
<sup>1</sup> Using the published genome size of 758 Mb			

# EL10 Assembly: outputs

Assembly by input and method	Name	# Contigs:	% Scaffolded	Total size	N50	% >100 nt	# Scaffolds:	Total size	N50	%N	Coverage % <sup>1</sup>
				----- (x 1,000 nt) -----				----- (x 1,000 nt) -----			
RefBeet 1.2 (Dohm et al. 2014)	RefBeet	60,051	93.7	517,882	43.8	1.0	40,508	566,571	2,013	8.6	nd
EL10.1 PacBio	SBJ_80X	938	na	562,760	1,394.2	70.9	938	562,760	1,394	0.0	89.6
EL10.1 PacBio BioNano	SBJ_80X_BN	2,983	99.2	533,042	1,339.5	21.5	86	566,848	12,513	5.9	90.3
EL10.1 PacBio BioNano Hi-C	EL10.1	364	96.2	540,479	2,700.6	96.7	40	540,537	57,939	0.01	86.1
	Chromosome 1	47	100	58,076	2,421.0	100.0	1	58,086	na	0.02	9.2
	Chromosome 2	30	100	54,968	2,834.0	96.7	1	54,972	na	0.01	9.2
	Chromosome 3	22	100	54,096	3,727.5	100.0	1	54,100	na	0.01	8.6
	Chromosome 4	47	100	61,154	2,396.1	97.9	1	61,163	na	0.01	9.7
	Chromosome 5	30	100	59,218	3,579.3	93.3	1	59,225	na	0.01	9.4
	Chromosome 6	52	100	65,091	2,380.7	98.1	1	65,097	na	0.01	10.4
	Chromosome 7	40	100	57,345	2,831.3	95.0	1	57,354	na	0.02	9.1
	Chromosome 8	37	100	57,932	2,335.2	97.3	1	57,939	na	0.01	9.2
	Chromosome 9	28	100	52,176	2,381.7	100.0	1	52,180	na	0.01	8.3
	Un scaffolded	31	0	20,421	1,679.4	87.1	31	20,421	na	0.00	3.3

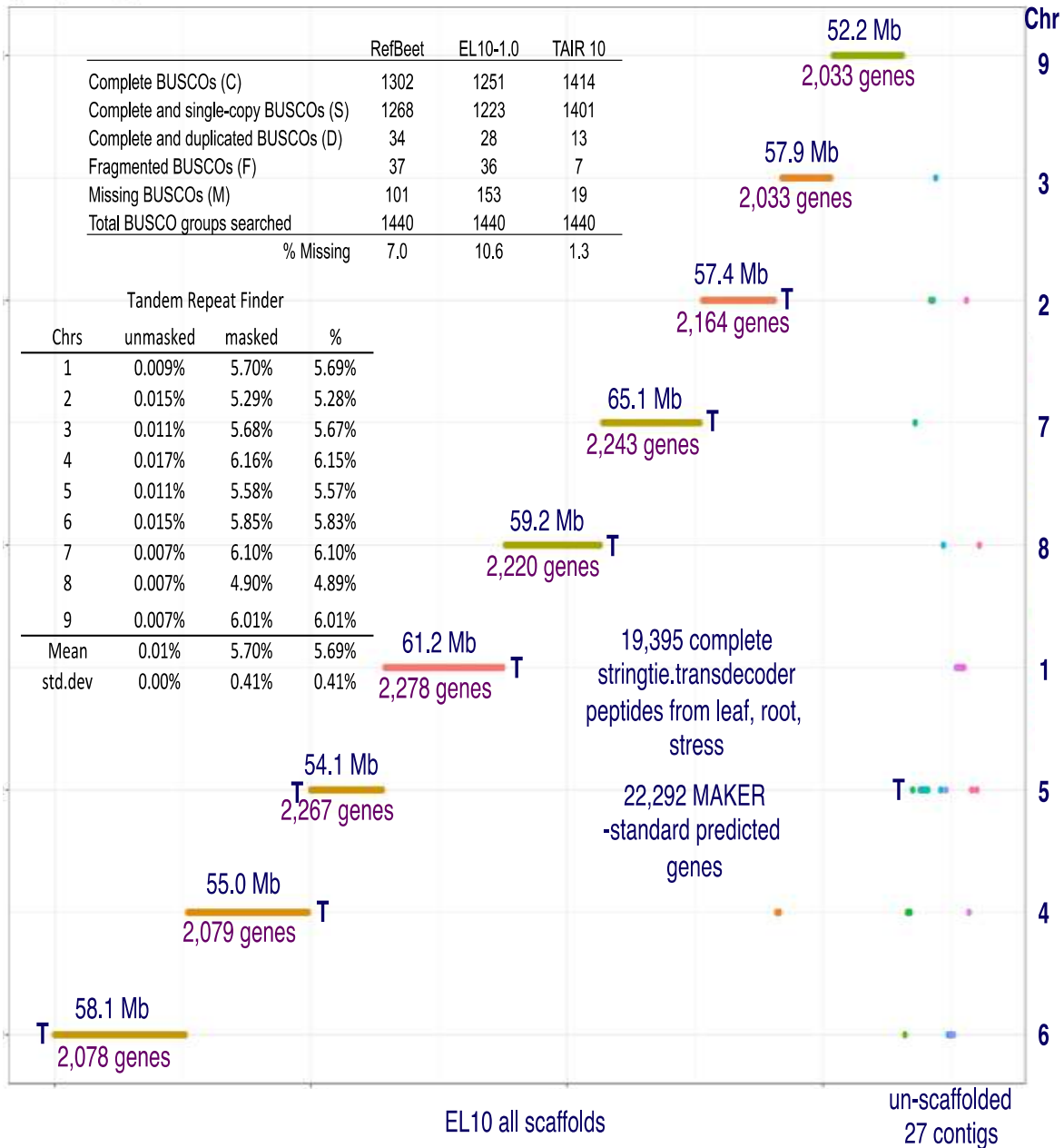
<sup>1</sup> Based on 628 Mb Physical Map

# Contiguity



EL10.1 Chromosomes

# Completeness



**EL10 is among the best plant assemblies created ...  
what can be done with it?**

Develop new genetic markers for breeding.

Correlate markers with traits.

**Examine occurrence and distribution of gene families.**

**Genome-wide evaluation of cultivars, breeding lines, and germplasm.**

Evaluate distribution of genetic diversity.

Predict genes involved in agronomic traits.

AT SINGLE NUCLEOTIDE RESOLUTION



# Rhizomania

Rhizomania resistance genes are located  
on Chromosome 3.

*Rz1* & *Rz2* are deployed commercially.

*Rz3*, *Rz4*, & *Rz5* are 'described'.

*Tz1* & *Rz3* is characterized as NB-ARCs.

NB-ARC domains are highly conserved.

How many NB-ARC domains are in EL10?



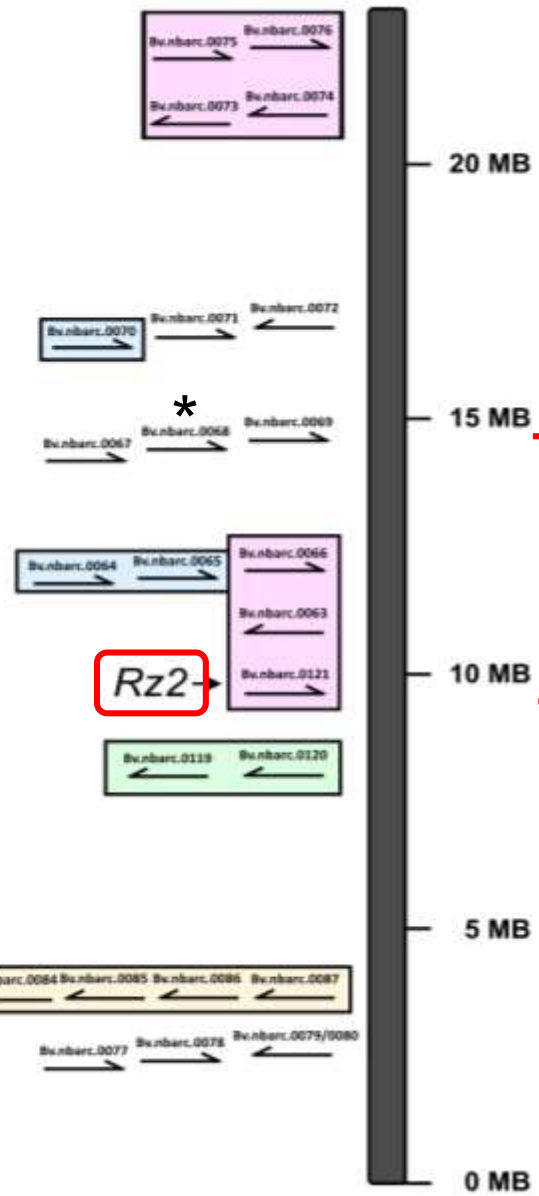
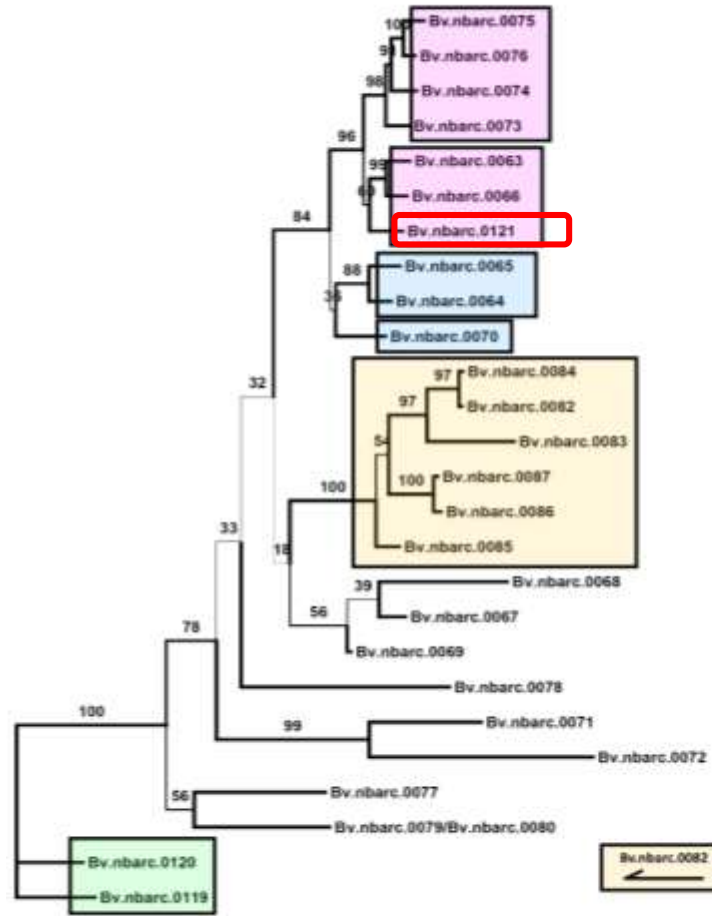


**Clades of NB-ARCs on Chromosome 3**

**Position of NB-ARCs on Chromosome 3**

**26 NB-ARCs**

~100 genes predicted in this ~ 20 Mb region



**Rz-region**

rhizomania resistance markers are located within the Rz-region

## **Gene discovery**

Breeding done with populations (most populations are variable).

Gene frequency estimates allow detection of past selection.

### **Approach :**

Illumina sequence 25 individuals to 80X total depth of genome coverage.

Map reads to genome(s).

Determine polymorphic sites (~ 14 million SNPs).

Filter to bi-allelic SNPs.

Calculate  $F_{ST}$  and  $2p_q$ .

Plot values across the genome assembly.

Interpret.

Crop type genes:  
22 populations examined.  
9 EL sugar, 7 table, 4 chard, 2 fodder.



Sugar

Chard

Fodder

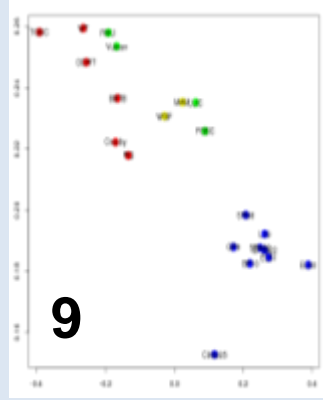
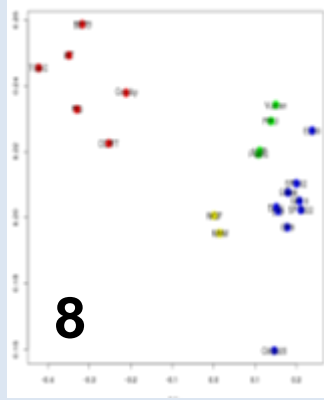
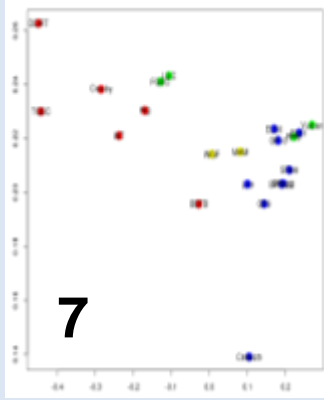
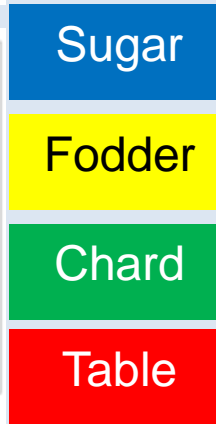
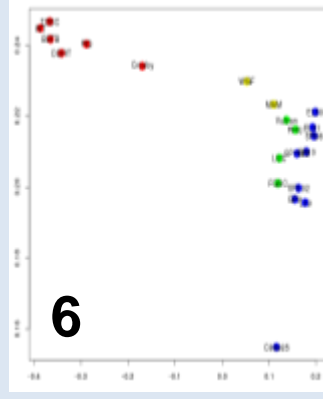
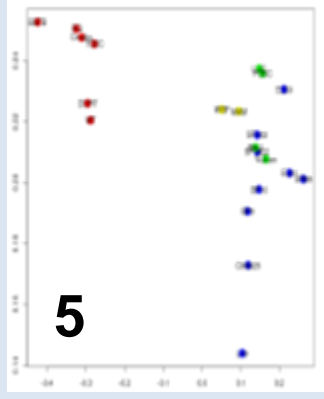
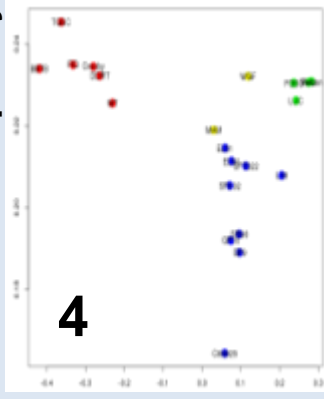
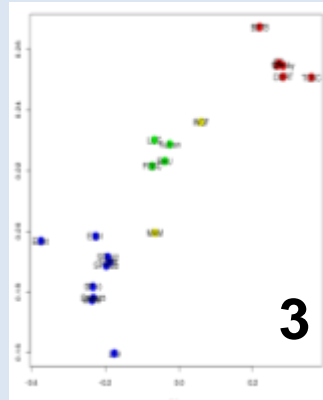
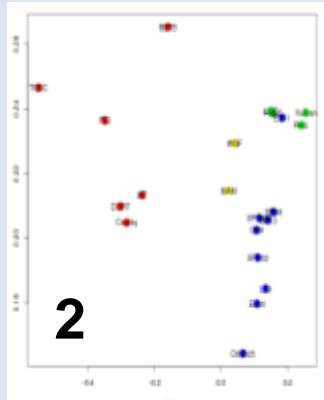
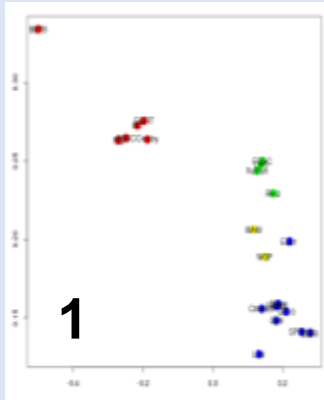
Table

9 week old plants

by Chromosome

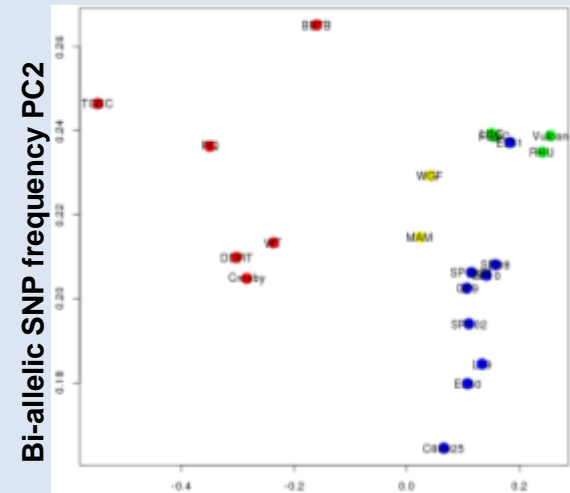
# Bi-allelic SNP frequency principal components

Bi-allelic SNP frequency PC2



Bi-allelic SNP frequency PC1

Genome-wide



Bi-allelic SNP frequency PC1



# Population genomics of Chromosome 9

4 crop types, 2-9 accessions, 25 plants per accession, 80X PE Illumina sequences

## $F_{ST}$

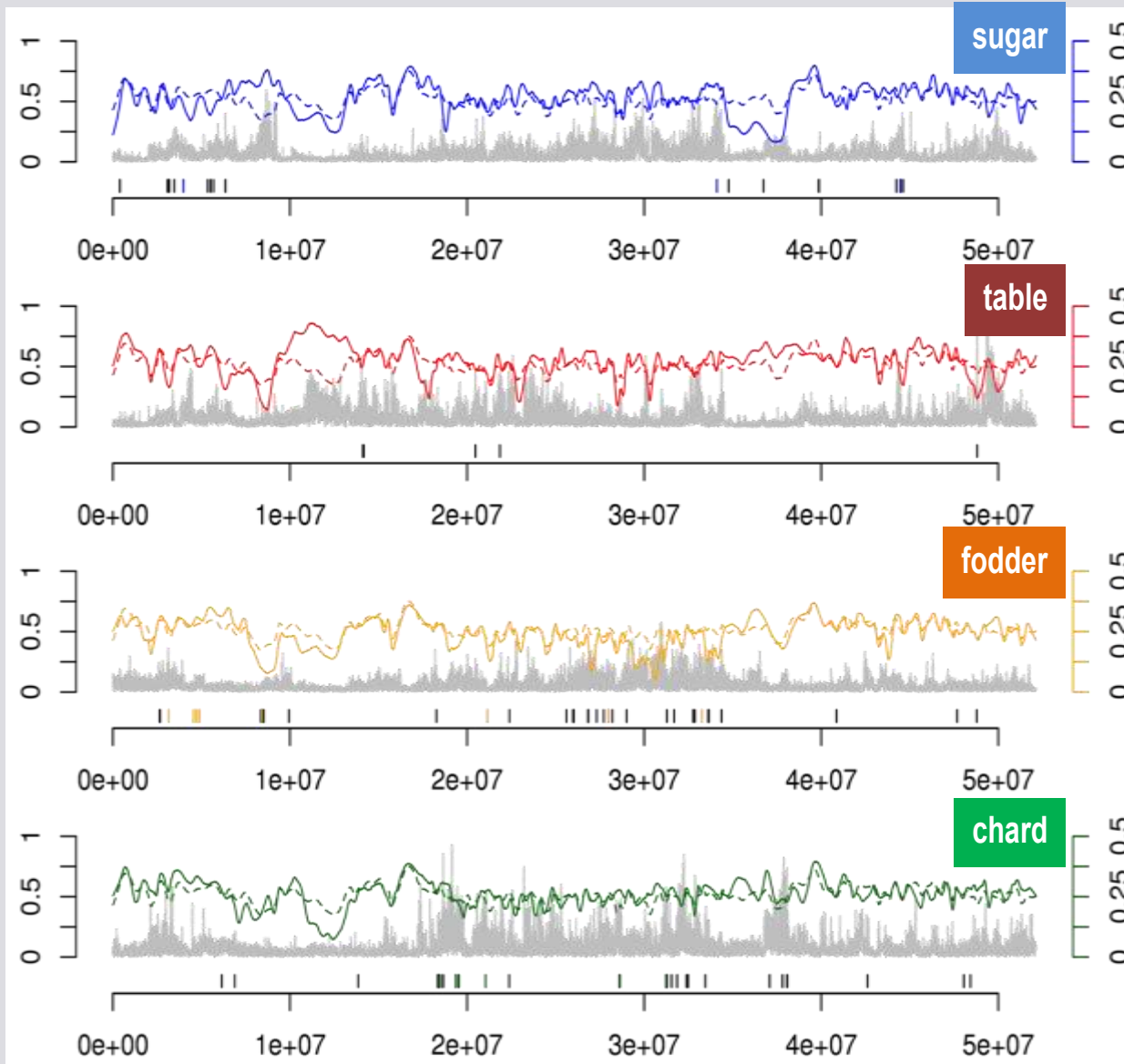
(histogram)

$F_{ST}$  fixation index,  
~ probability genes are  
identical by descent

uses average nucleotide  
differences

$$\frac{(\text{between pops} - \text{within pops})}{\text{between pops}}$$

ranges between 0 and 1



## $2pq$

(lines)

(dashed line is all  
populations averaged)

ranges between 0 and 0.5

$2pq$  = heterozygosity

$p^2 + 2pq + q^2 = 1$

nucleotide position



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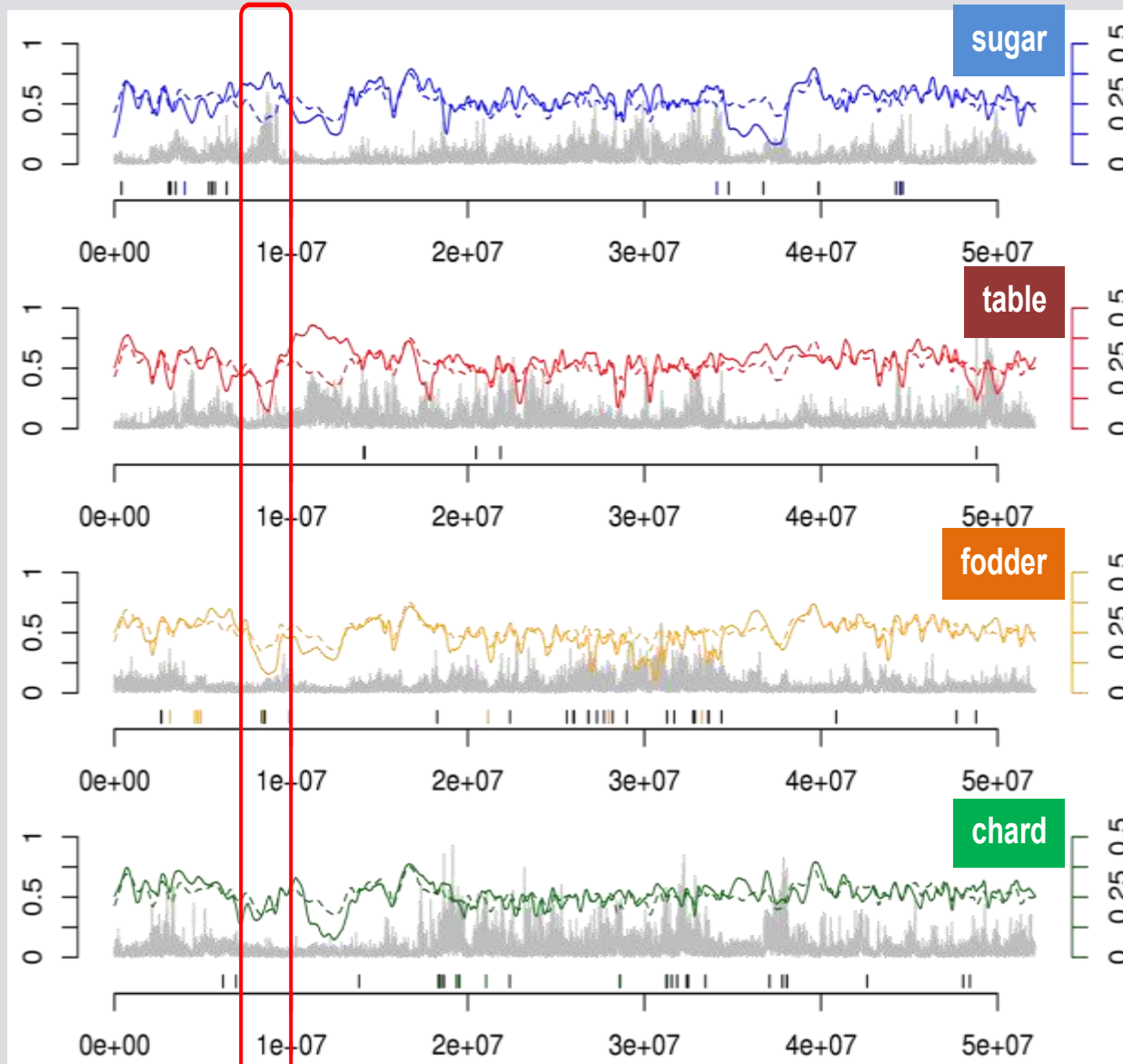
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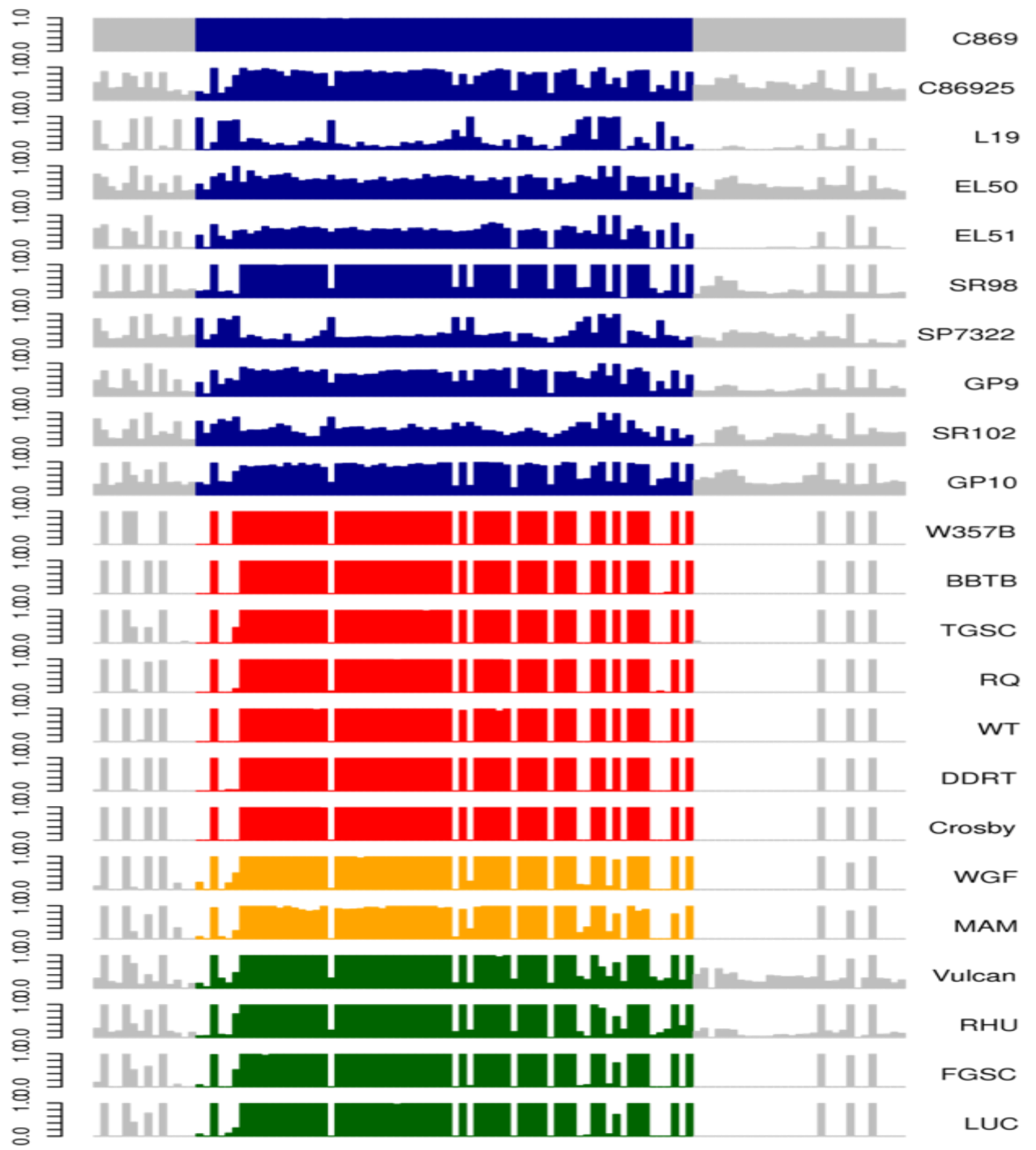
nucleotide position

# Zoom in on one gene in one FST peak of Chromosome 9

colored = coding  
grey = non-coding

$F_{ST}$

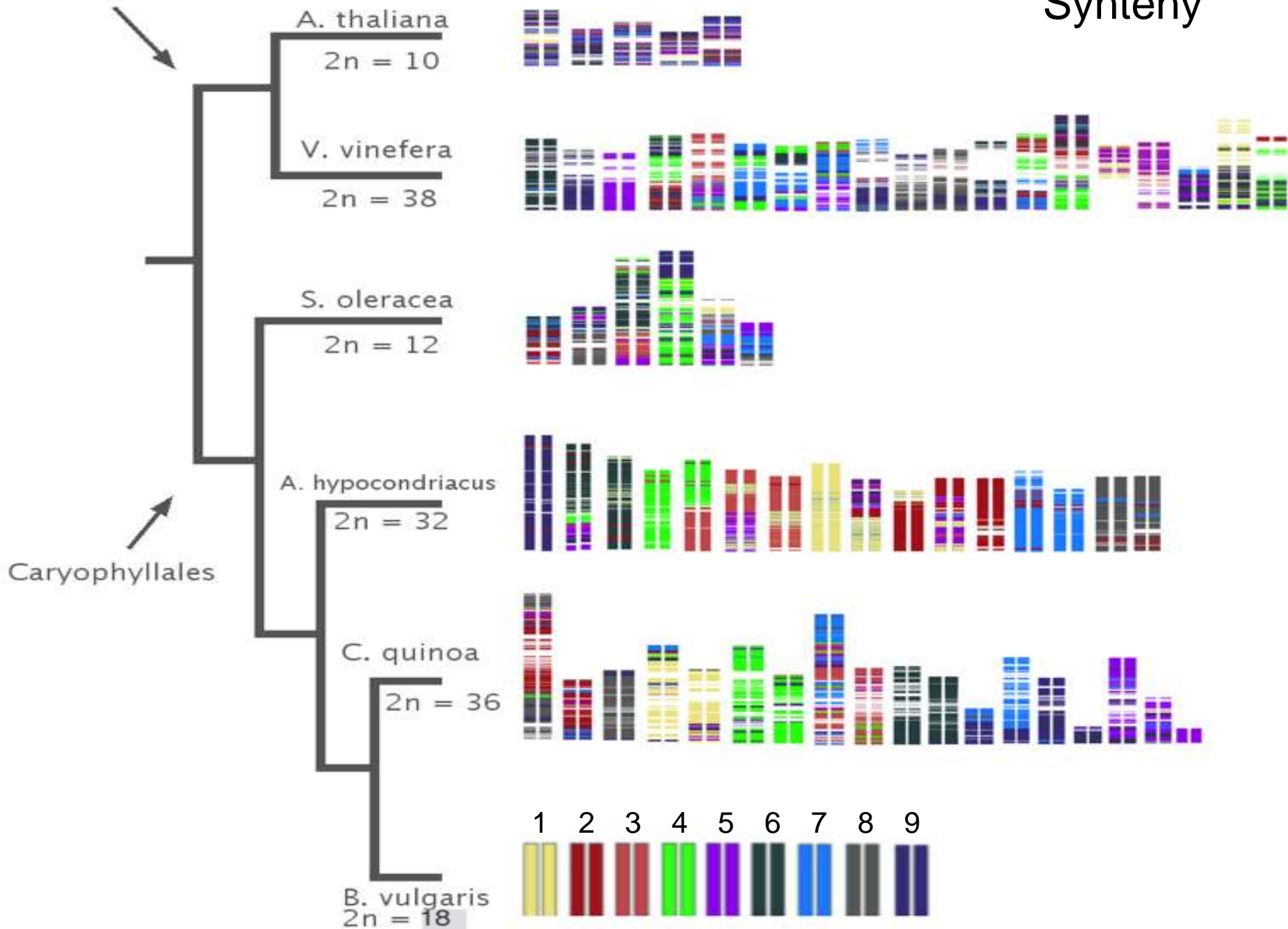
Use to develop testable hypotheses,  
~ 5,000 crop-wide FST peaks



CULTIVAR

Rosids

Synteny



*A. thaliana*

$2n = 10$

*V. vinefera*

$2n = 38$

*S. oleracea*

$2n = 12$

*A. hypochondriacus*

$2n = 32$

*C. quinoa*

$2n = 36$

*B. vulgaris*

$2n = 18$

1

2

3

4

5

6

7

8

9

## **Conclusion**

High-quality genome assembly.

Comprehensive targeting of specific genes and gene families of interest.

Genome-wide assessment of differentiation possible.

Single nucleotide resolution allows specific genetic hypotheses to be developed and tested.

**Thank you for your comments, questions, and your attention!**

